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Supplemental Material

Epigenome-Wide Meta-Analysis of Methylation in Children Related to Prenatal NO₂ Air Pollution Exposure

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Acknowledgments

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Materials and Methods

MeDALL

MeDALL (Mechanisms of the Development of ALLergy) is a collaborative project supported by the European Union under the Health Cooperation Work Programme of the 7th Framework programme (grant agreement number 261357) (Bousquet et al. 2011). MeDALL epigenetics studies include four birth cohorts. These are EDEN, BAMSE, PIAMA and INMA.

MeDALL – EDEN

The EDEN (Etude des Déterminants pré et post natals du développement et de la santé de l'Enfant) study is a prospective Birth Cohort Study (https://eden.vjf.inserm.fr/), which has been described in detail elsewhere (Heude et al. 2015). Pregnant women seen for a prenatal visit at the departments of Obstetrics and Gynecology of the University Hospital of Nancy and Poitiers before their twenty-fourth week of amenorrhea were invited to participate. Enrolment started in February 2003 in Poitiers and September 2003 in Nancy; it lasted 27 months in each centre. Among eligible women, 55% (2002 women) accepted to participate. The study has been approved by the ethical committees « Comité Consultatif pour la Protection des Personnes dans la Recherche Biomédicale », Le Kremlin-Bicêtre University hospital, and « Commission Nationale de l'Informatique et des Libertés ».

MeDALL - INMA

The INMA—INfancia y Medio Ambiente—(Environment and Childhood) Project is a network of birth cohorts in Spain that aim to study the role of environmental pollutants in air, water and diet during pregnancy and early childhood in relation to child growth and development (Guxens et al. 2012). The study has been approved by Ethical Committee of each participating centre and written consent was obtained from participating parents. Data for this study came from INMA Sabadell cohort (children born between 2004 and 2007). A total of 187 mothers with prenatal air pollution information and offspring cord blood DNA methylation were included, as well as 195 samples at age 4 years.

MeDALL - PIAMA

For the PIAMA birth cohort study, pregnant women were recruited in 1996-1997 during their second trimester of pregnancy from a series of communities in the North, West, and Centre of The Netherlands as described elsewhere (Wijga et al. IJE 2014). Non-allergic pregnant women were invited to participate in a "natural history" study arm. Pregnant women identified as allergic through a validated screening questionnaire were primarily allocated to

an intervention arm with a random subset allocated to the natural history arm. The intervention involved the use of mite-impermeable mattress and pillow covers. The study started with 3,963 newborns. Information on the children's health, socio-demographic and lifestyle factors as well as residential characteristics was collected by questionnaire annually until age 8 years and then at ages 11 and 14 years.

MeDALL - BAMSE

BAMSE is a prospective population-based cohort study of children recruited at birth and followed during childhood and adolescence. Details of the study design, inclusion criteria, enrolment and data collection are described elsewhere (Thacher et al. 2016; Wickman et al. 2002). In short, 4,089 children born between 1994 and 1996 in four municipalities of Stockholm County were enrolled. At baseline, when the infant was approximately 2 months of age, parents completed a questionnaire that assessed residential characteristics, as well as socioeconomic and lifestyle factors. When children were 1, 2, 4, 8, 12 and 16 years, the parents completed questionnaires focusing on children's symptoms related to wheezing and allergic diseases, as well as various exposures. The survey response rates were 96%, 94%, 91%, 84%, 82% and 78%, respectively. Furthermore, blood was obtained at ages 4, 8 and 16 years from 2,605 (63.7%), 2,470 (60.4%) and 2,547 (62.2%) children, respectively. The baseline and follow-up studies were approved by the Regional Ethical Review Board, Karolinska Institutet, Stockholm, Sweden, and the parents of all participating children provided informed consent.

MeDALL DNA methylation measurements

In the MeDALL study, peripheral blood samples were collected from all consenting cohort participants, and DNA from peripheral and cord blood samples was extracted using the QIAamp blood kit (Qiagen, Inc, Valencia, CA) or equivalent protocols, followed by a precipitation-based concentration using GlycoBlue (Ambion, Austin, Tex). DNA concentration was determined by Nanodrop measurement and picogreen quantification. 500 ng of DNA were bisulfite-converted using the EZ 96-DNA methylation kit following the manufacturer's standard protocol, and DNA methylation measured using the Illumina Infinium HumanMethylation450 beadchip (Illumina, Inc., San Diego, USA). DNA methylation data were preprocessed using the Minfi package (Aryee et al. 2014).

In quality control, samples that did not provide significant methylation signals in more than 10% of probes (detection p-value=0.01) were regarded as bad quality samples and were directly removed. In addition, samples were excluded in case of low staining efficiency, low single base extension efficiency, low stripping efficiency of DNA from probes after single base extension, poor hybridization performance, poor bisulfite conversion and high negative control probe staining. Moreover, we used 65 SNP probes to check for concordances between paired DNA samples from the sample individual and assessed the methylation distribution of X-chromosome to verify gender. Paired samples which show Pearson correlation coefficient <0.9 were regarded as sample mixed ups and were excluded from the study. Furthermore, we excluded probes on sex chromosomes, probes that mapped on multi-loci, the 65 random SNPs assay, and probes that contained SNP(s) at the target CpG sites with a minor allele frequency >10%. A series of steps were completed for quality control and data analysis. First, we implemented sample filtering to remove bad quality and mixed up samples. Second, we filtered out the probes to remove the CpG sites which are not mapped to unique location on the genome and CpGs containing single nucleotide polymorphisms (SNPs) at the target site. Third, we implemented "DASEN" (Pidsley et al. 2013) to perform signal correction and normalization.

To remove bias in methylation profiles unrelated to underlying biological processes, we implemented a correction procedures based on 613 negative control probes presented in 450K arrays since these negative control probes are supposed to not relate to biological variation. Finally, we implemented principal component analysis (PCA) on control probes data, then, we performed 10000 permutation for controls probes data and selected principal components with p-value defined as to get the p-value of (number of var(random pc) >var(pc)) / (number of permutations) <10⁻⁴. The methylation data for each CpG are thus the residuals from a linear model fitting incorporating the significant 5 PCs.

MeDALL gene expression measurements

Data on mRNA gene expression were available in the BAMSE (239 children aged 16 years) and the INMA (111 children aged 4 years) cohorts. Whole blood was collected in PAXGene tubes and RNA was extracted using PAXgene Blood RNA kit (QIAGEN, Courtaboeuf, France). Quantity of extracted RNA and quality assessment were performed with Dropsense96 (Trinean, Gentbrugge, Belgium) and Tapestation (Agilent, Les Ulis, France) instruments, respectively, discarding 3 samples (BAMSE n=2, INMA n=1). RNA of

highest quality was selected for amplification, labeling and hybridization on Affymetrix HTA 2.0 Genechips using Affymetrix IVT kit (Affymetrix, Inc. USA) at the European Institute for Systems Biology and Medicine in Lyon. Data were then processed at the probesets level for RMA normalization using Expression Console Software from Affymetrix v1.4. Expression transcripts were annotated using version 35 of Affymetrix annotation. In the INMA cohort, 12 samples were excluded because they were outliers defined as more than 3SD from the mean for PC1 or PC2 (n=8) or there were sex discrepancies (n=8). The Affy HTA 2.0 technology we used is composed of 428, 250, 328, 242, 80 and 126 probes for the *TPO, CAT, LONP1*, *SLC25A28, PLVAP* and *GPR55*, respectively. Each of these sets contains probes that are scattered at various localization along the transcripts, including exons and junctions, averaged into one Transcript Cluster (per gene) summarizing the specific gene expression level. For the BAMSE samples, automated cell count was obtained by flow cytometry performed at the Karolinska University Laboratory in Stockholm, Sweden.

BAMSE EpiGene

In the BAMSE cohort we also had an independent methylation dataset. Epigenome-wide DNA methylation was measured in 472 Caucasian children, using DNA extracted from blood samples collected at the age of 8 years. An aliquot (500 ng) of DNA per sample underwent bisulfite conversion using the EZ-96 DNA Methylation kit (Zymo Research Corporation, Irvine, USA). Samples were plated onto 96-well plates in randomized order. Samples were processed with the Illumina Infinium HumanMethylation450 BeadChip (Illumina Inc., San Diego, USA).

Quality control of analysed samples was performed using standardized criteria. Samples were excluded in case of sample call rate <99%, colour balance >3, low staining efficiency, poor extension efficiency, poor hybridization performance, low stripping efficiency after extension and poor bisulfite conversion. We also applied multidimensional scaling (MDS) plot to evaluate gender outliers based on chromosome X data, that produced two separated clusters for male and female. We omitted 5 samples that do not belong to the distinct cluster. Furthermore, we applied median intensity plot for methylated and unmethylated intensity by using the minfi R package (3 samples below the 10.5 cutoff were excluded). All above led to exclusion of 8 samples.

Probes with a single nucleotide polymorphism in the single base extension site with a frequency of >5% were excluded (Chen et al. 2013), as were probes with non-optimal binding (non-mapping or mapping multiple times to either the normal or the bisulphite-converted genome), and the probed belonging to chrX and ChrY, resulting in the exclusion of 46,799 probes, leaving a total of 438,713 probes in the analysis.

Furthermore, we implemented "DASEN" recommended from wateRmelon package to do signal correction and normalization (Pidsley et al. 2013).

The Generation R Study

The Generation R Study is a population-based prospective cohort study from fetal life onwards in Rotterdam, the Netherlands, which has been previously described in detail (Jaddoe et al. 2012). Assessments in pregnant women consisted of physical examinations, fetal ultrasounds, biological samples, and questionnaires (Kruithof et al. 2014). All children were born between April 2002 and January 2006. The study has been approved by the Medical Ethical Committee of the Erasmus University Medical Centre and written consent was obtained for all participating mothers and children. For the current study, data was available for 809 Caucasian mothers and their children with information on NO₂ exposure during pregnancy and DNA-methylation at birth.

Generation R DNA methylation measurements

DNA was extracted from cord blood samples of 979 Caucasian children. Using the EZ-96 DNA Methylation kit (Shallow-well, Zymo Research Corporation, Irvine, USA), 500 ng DNA per sample underwent bisulfite conversion. Samples were transferred onto 96-well plates in a random order. Samples were processed with Illumina's Infinium HumanMethylation450 BeadChip (Illumina Inc., San Diego, USA). Quality control of analyzed samples was performed using standardized criteria. Samples were excluded due to sample call rate <99% (n=7) or poor bisulfite conversion (n=1). In addition, 2 samples were excluded because of a gender mismatch and 1 sample because of a retracted informed consent, leaving a total of 969 samples in the statistical analysis.

Probes with a single nucleotide polymorphism in the single base extension site with a frequency of >1% in the GoNLv4 reference panel were excluded, as were probes with non-optimal binding (non-mapping or mapping multiple times to either the normal or the

bisulphite-converted genome), resulting in the exclusion of 49,564 probes, leaving a total of 436,013 probes in the analysis.

Data were normalized with DASES normalization using a pipeline adapted from that developed by Touleimat and Tost (Touleimat and Tost 2012). DASES normalization includes background adjustment, between-array normalization applied to type I and type II probes separately, and dye bias correction applied to type I and type II probes separately. DASES is based on the DASEN method, but adds the dye bias correction, which is not included in DASEN (Pidsley et al. 2013). Beta-values were calculated for all CpG sites.

Air pollution exposure assessment in MeDALL, the Generation R Study and BAMSE EpiGene

The procedures for measurements and LUR modeling have been extensively described elsewhere (Pedersen et al. 2013; Van den Hooven et al. 2012). In short, 40 sampling sites (80 in the Netherlands/Belgium) for NO₂ and other agents were selected in each study area to characterize the spatial distribution of the cohort addresses, including regional background, urban background, and traffic sites. Measurements were performed at each site 3 times during 2 weeks in the cold, warm, and intermediate seasons, and the results were averaged to estimate the annual average. LUR models for NO₂ were developed based on measured annual average concentrations by using a range of Geographic Information System—derived predictor variables selected through a supervised stepwise procedure. Modeling was done locally at each center according to a common exposure assessment manual (http://www.escapeproject.eu/manuals/) following harmonized procedures regarding air pollutants measurements, development of land use regression models, and validation (Beelen et al. 2013). For the present analyses, total NO₂ exposure averaged throughout entire pregnancy was used. Data from routine monitoring stations were used to temporally adjust the LUR estimates to the periods corresponding to each individual pregnancy. The current

Children's Health Study (CHS)

CHS is a population-based prospective cohort study from age 6 onwards in Southern California, which has been described in detail elsewhere (McConnell et al. 2006). The study protocol was approved by the University of Southern California Institutional Review Board

exposure was estimated by assignment of modeled annual average NO₂ concentrations to the

current addresses at the time when the blood samples were collected.

and informed, written consent and assent were provided by the parents and children respectively.

A total of 5,341 children were recruited from schools within several Southern California communities, all of whom were born between 1995 and 1997 and are currently being followed until age 18. The cohort was established with the purpose of investigating the effects of air pollutants on respiratory health in children. Personal, parental, and sociodemographic characteristics were obtained by parent-completed questionnaire. Birth weight, gestational age, mode of delivery and other reproductive data were obtained from California birth records. The estimated date of conception was assigned using the birth date and gestational age, corrected for the average 2-week difference between the last menstrual period and conception. Ancestry (European, African, and Asian) was measured using ancestry informative markers SNPs and included as an additional covariate.

CHS DNA methylation measurements

Epigenome-wide DNA methylation was measured in 226 Hispanic and non-Hispanic white children, using DNA extracted from newborn bloodspots archived by the state of California. Laboratory personnel performing DNA methylation analysis were blinded to study subject information. DNA was extracted whole blood cells using the QiaAmp DNA blood kit (Qiagen Inc, Valencia, CA) and stored at -80 degrees Celcius. 700-1000ng of genomic DNA from each sample was treated with bisulfite using the EZ-96 DNA Methylation KitTM (Zymo Research, Irvine, CA, USA), according to the manufacturer's recommended protocol and eluted in 18 ul. The results of the Infinium HumanMethylation450 BeadChip (HM450) were compiled for each locus as previously described and were reported as beta (β) values (Noushmehr et al. 2010).

CpG loci on the HM450 array were removed from analyses if they were on the X and Y chromosomes, or if they contained SNPs, deletions, repeats, or if they have more than 10% missing values. Data were processed in the methylumi package in R, after which a normal exponential background correction was applied to the raw intensities at the array level to reduce background noise (Triche et al. 2013). We then normalized each sample's methylation values to have the same quantiles to address sample to sample variability (Bolstad et al. 2003). β-values were calculated for all CpG sites.

CHS air pollution exposure assessment

The CHS air quality monitoring data (Peters et al. 1999a; Peters et al. 1999b) and the US EPA air Quality System were used to assign estimates of prenatal air pollution exposures for NO₂, based on a combination of residential history obtained from parents when subjects were 6-7 years old and birth address recorded on the birth certificate. In all but 34 cases, questionnaire-reported birth address from the residential history matched the birth address from the birth certificate. In the 34 cases where a mismatch was identified, the birth certificate address was used to assign air pollution exposure. Moreover, the birth address was representative of the mother's location throughout pregnancy in 88% of the subjects.

Addresses were geocoded using TeleAtlas Inc.'s Address Point Geocoding Services. Station-specific air quality data were spatially interpolated to each birth residence using inverse-distance-squared weighting (Hannam et al. 2013; Rivera-Gonzalez et al. 2015). The data from up to four air quality measurement stations were included in each interpolation with a maximum interpolation radius of 50 km. However, when a residence was located within 5 km of one or more stations with valid observations, the interpolation was based solely on the nearby values. Prenatal ambient air pollution concentrations were estimated for each subject's reported birth residence based on average monthly air pollutant exposure data. A leave one out evaluation of the spatial mapping method produced an r²=0.73, for monthly NO₂ concentrations using data from California.

The Norwegian Mother and Child Cohort (MoBa)

MoBa is a prospective population-based pregnancy cohort study conducted by the Norwegian Institute of Public Health (Magnus et al. 2006; Ronningen et al. 2006). Participants were recruited from all over Norway from 1999-2008. The women consented to participation in 40.6% of the pregnancies. The cohort includes about 114,500 children, 95,200 mothers and 75,200 fathers. Blood samples were obtained from both parents during pregnancy and from mothers and children (umbilical cord) at birth. Data from MoBa was linked to the Medical Birth Registry of Norway (MBRN), and the current study used information from follow-up questionnaires at approximately 18-, and 30 - gestational weeks. DNA methylation and information on NO₂ exposure were available on 193 pregnancies. MoBa has obtained a licence from the Norwegian Data Inspectorate, and the current study was approved by The Regional Committee for Medical Research Ethics

MoBa DNA methylation measurements

Details on the DNA methylation and quality control can be found in (Joubert et al. 2016).

MoBa air pollution exposure assessment

Estimating air pollution exposure during pregnancy (including NO₂ measurements, LUR models development, and validation) was based on the methodology developed for the ESCAPE project (Beelen et al. 2013; Pedersen et al. 2013). LUR models for NO₂ levels were built for the studied areas in order to account for regional specifics. In the models we used air pollution measurements conducted in 2010 for Oslo and Akershus, and in 2011 for Bergen and Hordaland. Measurement campaigns included three rounds of approximately two weeks duration with NO₂ measurements (during winter, summer and an intermediate season) within a one year period. Measurement sites were selected to represent the range of residential exposure for each study area. For the analysis we included sites with no missing data, and no geocoding mismatches. LUR-modelled NO₂ yearly mean estimates for residential addresses at birth were temporally adjusted for each individual pregnancy using continuous routine monitoring station data. Daily NO₂ exposure estimates were averaged over the whole pregnancy.

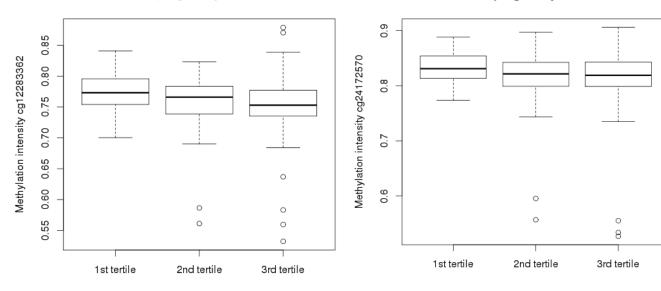
Table S1. Characteristics of the included cohorts with available cord blood samples

			MeDAI	LL cohorts			
Cohort characteristics	BAMSE	EDEN	INMA	PIAMA	CHS	Generation R	MoBa
Study design	Population based birth cohort.	Population based birth cohort, enrolled before 26 weeks of pregnancy	Population based birth cohort, enrolled at week 12 of pregnancy	Population based birth cohort (with mattress cover intervention and allergic/non- allergic parents)	Population-based children's cohort with retrospective collection of residential history, and archived newborn bloodspots	Population-based prospective birth cohort	Population based birth cohort
Age at enrolment	Newborns	Pregnant women		Pregnant women	5-6 years	Pregnant women	Pregnant women
Population source (area)	Stockholm, Sweden	Nancy and Poitiers, France	Sabadell, Spain	North, West and center of the Netherlands	Greater Los Angeles area	Rotterdam, the Netherlands	Norway
Enrolment period	1994-1996	2003-2006	2004-2007	1996-1997	2002	2002-2006	1999-2008
Cohort recruitment	Community population register	Prenatal Healthcare	Prenatal healthcare	Prenatal healthcare	Community schools	Prenatal healthcare	Prenatal healthcare
Total number of recruited children	4,089	2,002	638	3,963	5341	9,901	
Follow-up time points (year of life)	1,2,4,8	Birth, 4 and 8 months, 1, 2, 3, 4, 5-6, 8 years	Birth, 1, 2, 4, 7, 9	1,2,3,4,5,6,7,8, 11,14	6-18		Birth, 0.5, 1.5, 3, 5, 7
Traffic air pollution							
Estimation model	LUR	LUR	LUR	LUR	Dispersion	LUR	LUR
Time of measurements	2009	2002, 2005	2010	2009	1995-1996	2002-2006	2010, 2011
Sample selection							
Criteria for selection of sample	4yr – 8yr paired DNA samples. Asthma case-control	Birth-5yr paired DNA samples.	Birth-4yr paired DNA samples. Asthma case-control	4yr – 8yr paired DNA samples. Asthma case-control	Randomly selected from within subjects that matched to CA birth records and had a bloodspot	European ancestry	

Methylation measurement platform	Illumina 450K assay	Illumina 450K assay	Illumina 450K assay	Illumina 450K assay	Illumina 450K assay	Illumina 450K assay	Illumina 450K assay
Number of CpGs available per cohort	439,306	439,306	439,306	439,306	368,386	436,013	473,731
Ethnicity	Caucasian	Caucasian	Caucasian	Caucasian	Hispanic white and non-Hispanic white	Caucasian	Caucasian

NO2 pregnancy P=0.001

NO2 pregnancy P=0.004



C) cg08973675 (SLC25A28)

NO2 pregnancy P=0.015

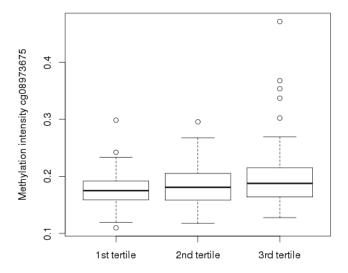


Figure S1. Plot of the cumulative distribution function for methylation intensities for the three top-ranked CpG sites: cg12283362 (*LONP1*), cg24172570 (3.8 kbp upstream of *HIBADH*), and cg08973675 (*SLC25A28*) by the cohort-specific tertiles of prenatal NO₂ exposure (in the INMA cohort, 1st tertile <39.9 μ g/m³; 2nd tertile 39.9-47.6 μ g/m³; 3rd tertile >47.6 μ g/m³; in the EDEN cohort, 1st tertile <12.7 μ g/m³; 2nd tertile 12.7-17.6 μ g/m³; 3rd tertile >17.6 μ g/m³) demonstrates a statistically significant dose-response effect of exposure in the INMA and EDEN cohorts. The p-values are derived from the Jonckheere-Terpstra trend test, calculated using the SAGx package in R, version 2.14.0.

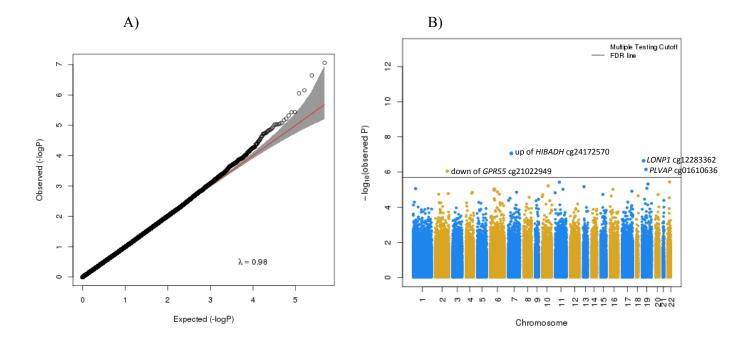


Figure S2. Quantile-quantile plot (A) and Manhattan plot (B) for epigenome-wide metaanalysis of the association between NO₂ exposure during pregnancy and cord blood DNA methylation, additionally adjusted for cell type composition (n=1,508). Four CpGs were considered statistically significant using FDR correction (solid horizontal line): cg24172570 3.8 kbp upstream of *HIBADH*, cg12283362 in *LONP1*, cg01610636 in *PLVAP*, and cg21022949 19.7 kbp downstream of *GPR55*.

Table S2. Top 25 CpGs from the epigenome-wide meta-analysis of the association between NO₂ exposure during pregnancy and cord blood DNA methylation additionally adjusted for cell composition (n=1,508 newborns from MeDALL, Generation R, CHS and MoBa cohorts).

Chr	Position (build 37)	CpG	Mapped gene	Gene group	Coef	SE	P-value	Direction
7	27561178	cg24172570	HIBADH* FDR		-0.004	8.00E-04	8.65E-08	?-
19	5709149	cg12283362	LONP1 FDR	Body	-0.007	1.30E-03	2.27E-07	-??-
19	17463255	cg01610636	PLVAP FDR	Body	-0.005	9.00E-04	7.03E-07	
2	231809697	cg21022949	$GPR55*^{FDR}$		0.001	2.00E-04	8.86E-07	++++
22	40355732	cg17988310	GRAP2	Body	0.004	8.00E-04	3.65E-06	++++
11	47400146	cg03565868	SPI1	TSS200	0.005	1.10E-03	3.71E-06	+++-
19	39884218	cg14651844	MED29	Body	0.002	5.00E-04	4.75E-06	+-++
10	101380289	cg08973675	SLC25A28	TSS200	0.005	1.10E-03	6.05E-06	+++-
13	32524761	cg00648883	EEF1DP3	Body	0.006	1.30E-03	6.77E-06	++++
19	30155866	cg05512099	PLEKHF1	TSS1500	-0.007	1.60E-03	8.36E-06	-+?-
1	9675560	cg25590444	TMEM201*		0.003	6.00E-04	8.67E-06	+-++
6	31382102	cg26504614	MICA	Body	-0.005	1.10E-03	9.25E-06	-?
6	30524763	cg03860665	PRR3	5'UTR; 1stExon	0.002	4.00E-04	9.27E-06	++++
16	27325254	cg06641959	IL4R	5'UTR;1stExon	0.002	4.00E-04	9.38E-06	++++
11	74871202	cg12537437	SLCO2B1	Body;5'UTR	-0.003	7.00E-04	9.47E-06	+
6	30689865	cg25742745	TUBB	Body	0.004	1.00E-03	1.09E-05	+??+
17	78926091	cg21831512	RPTOR	Body	0.004	8.00E-04	1.22E-05	++
6	33359817	cg04757012	KIFC1	Body	0.001	3.00E-04	1.33E-05	++++
7	117824040	cg08301459	NAA38	TSS200	0.002	3.00E-04	1.39E-05	++?+
4	154386136	cg21908828	KIAA0922	TSS1500	0.004	1.00E-03	1.42E-05	++++
11	369155	cg19787465	B4GALNT4	TSS1500	0.004	8.00E-04	1.49E-05	+++-
4	139940407	cg04702527	CCRN4L	Body	-0.005	1.20E-03	1.58E-05	-+?-
2	239984105	cg09155776	HDAC4	Body	-0.010	2.30E-03	1.69E-05	
2	60698937	cg04588436	BCL11A	Body	-0.003	8.00E-04	1.80E-05	-??-
6	56114591	cg11241549			0.007	1.50E-03	1.81E-05	+??+

Shown are top 25 CpGs ordered by p-value; All results presented per 10 μg/m³ increase in prenatal NO₂ exposure. Column headers: CHR: chromosome; Position: Chromosomal position based on NCBI human reference genome assembly Build 37. Mapped Gene: UCSC annotated gene; Gene Group: UCSC gene region feature category; regression coefficient; SE: standard error for regression coefficient; Direction: Direction of effect across cohorts included in the statistical model (MeDALL, Generation R, CHS and MoBa): NO₂ exposure during pregnancy associated with increased (+) or decreased (-) methylation, or missing (?) result. Genome-wide significance threshold (FDR p<0.05).

^{*}cg24172570 is located 3.8 kbp upstream of HIBADH; cg21022949 – 19.7 kbp downstream of GPR55; cg25590444 - 0,6 kbp downstream of TMEM201.

** Data on methylation of cg12283362 was available in 473 individuals, cg24172570 - in 1282 individuals.

Table S3. Top 25 CpGs from the analysis of the association between NO₂ exposure during pregnancy and cord blood DNA methylation without adjustment for cell composition (upper part) and with adjustment for cell composition according to Bakulski (lower part) (n=280 newborns from the MeDALL study).

	Unadjusted for celltype									
		NO ₂ at pregnai	$ncy \rightarrow Methylation at bi$		·INMA n	=280)				
	Position			Gene						
Chr	(build 37)	CpG	Mapped gene	group	Coef	SE	P-value			
17	43394547	cg03787849	MAP3K14 FDR	Island	0.002	4.09E-04	7.69E-08			
19	5709149	cg12283362	LONP1 FDR	S_Shore	-0.007	1.42E-03	3.62E-07			
11	62439187	cg26435734	C11orf83;C11orf48 FDR	_ Island	0.002	4.03E-04	5.72E-07			
12	121124951	cg26413987	$MLEC^{FDR}$	Island	0.002	3.34E-04	7.24E-07			
1	24127146	cg13180787	GALE FDR	Island	0.004	7.56E-04	1.15E-06			
19	14201991	cg17688733	SAMD1 FDR	OpenSea	0.003	5.89E-04	1.37E-06			
2	122494609	cg04002021	MKI67IP ^{FDR}	S_Shore	0.003	6.14E-04	1.46E-06			
14	74416831	cg19496328	FAM161B;COQ6 FDR	N Shore	0.002	5.21E-04	1.67E-06			
		-		_						
2	25896320	cg20347626	DTNB FDR	Island	0.002	3.65E-04	2.10E-06			
19	4066280	cg06403289	ZBTB7A FDR	Island	0.005	9.65E-04	2.16E-06			
6	33290736	cg07689821	DAXX	Island	0.005	9.54E-04	2.18E-06			
8	65711658	cg01510388	CYP7B1	Island	-0.011	2.36E-03	2.72E-06			
6	31869521	cg11801452	ZBTB12	Island	0.003	6.09E-04	2.85E-06			
7	27561178	cg24172570	HIBADH*	OpenSea	-0.005	1.05E-03	3.35E-06			
10	101380289	cg08973675	SLC25A28	Island	0.007	1.48E-03	3.47E-06			
3	52001995	cg07110217	PCBP4	OpenSea	0.002	5.18E-04	3.61E-06			
11	118307669	cg18705039	MLL	Island	0.009	1.92E-03	4.13E-06			
6	28351351	cg15964468	ZSCAN12	Island	0.009	2.04E-03	4.31E-06			
6	30899568	cg26672776	SFTA2	OpenSea	-0.014	3.13E-03	4.38E-06			
2	110873741	cg09113530	MALL	S_Shore	-0.007	1.52E-03	4.62E-06			
4	95679705	cg09156233	BMPR1B	Island	-0.001	1.87E-04	4.74E-06			
11	1968310	cg18482326	MRPL23	N_Shore	0.005	1.18E-03	5.44E-06			
19	39897430	cg07051257	ZFP36	Island	0.006	1.27E-03	5.68E-06			
12	7070317	cg22260508	PTPN6	OpenSea	0.004	8.60E-04	6.17E-06			
17	78851213	cg08314949	RPTOR	S_Shore	0.016	3.46E-03	6.29E-06			
		Adi	usted for celltype accord	ing to Raku	lski					
			according + constant = constant			=280)				
	Position	- 2 F8	<u> </u>	Gene		/				
Chr	(build 37)	CpG	Mapped gene	group	Coef	SE	P-value			
19	5709149	cg12283362	LONP1 FDR	S_Shore	-0.007	1.28E-03	1.37E-07			
2	122494609	cg04002021	MKI67IP ^{FDR}	S Shore	0.003	5.40E-04	1.71E-07			
6	43244304	cg04002021	TTBK1 ^{FDR}	Island	-0.003	1.68E-03	6.08E-07			
	43244304	-	MAP3K14 ^{FDR}							
17		cg03787849		Island	0.002	4.49E-04	6.08E-07			
17	42989137	cg20911989	GFAP FDR	Island	-0.009	1.72E-03	7.31E-07			
12	121124951	cg26413987	MLEC FDR	Island	0.002	3.29E-04	8.23E-07			
17	77768687	cg04986373	CBX8 FDR	N_Shore	0.006	1.31E-03	9.56E-07			
8	65711658	cg01510388	CYP7B1 FDR	Island	-0.011	2.24E-03	1.13E-06			
14	74416831	cg19496328	FAM161B;COQ6 FDR	N_Shore	0.002	5.01E-04	1.50E-06			

11	118307669	cg18705039	MLL FDR	Island	0.009	1.83E-03	1.54E-06
19	4066280	cg06403289	ZBTB7A FDR	Island	0.005	9.70E-04	1.57E-06
2	241498221	cg24958325	ANKMY1;DUSP28 FDR	N_Shore	0.004	7.30E-04	1.58E-06
6	30899568	cg26672776	SFTA2 FDR	OpenSea	-0.014	2.98E-03	1.72E-06
3	52443970	cg10956904	BAP1;PHF7 FDR	Island	0.003	5.35E-04	1.89E-06
16	75021994	cg06383207		S_Shelf	-0.007	1.56E-03	2.41E-06
20	3030247	cg25472862		S_Shelf	0.003	7.39E-04	2.46E-06
10	101380289	cg08973675	SLC25A28	Island	0.007	1.45E-03	2.71E-06
19	14201991	cg17688733	SAMD1	OpenSea	0.003	5.60E-04	2.88E-06
6	33393786	cg21726235	SYNGAP1	Island	0.004	8.75E-04	2.95E-06
3	52001995	cg07110217	PCBP4	OpenSea	0.002	5.32E-04	3.44E-06
8	48099615	cg03271173		N_Shore	-0.003	5.87E-04	3.88E-06
1	1821981	cg03716942	GNB1	Island	0.002	4.87E-04	4.56E-06
16	30106682	cg26709300	YPEL3	N_Shore	0.006	1.41E-03	5.11E-06
19	17463255	cg01610636	PLVAP	S_Shore	-0.005	1.13E-03	5.16E-06
11	62439187	cg26435734	C11orf83;C11orf48	Island	0.002	4.22E-04	5.30E-06

Estimates are presented per 10 μg/m³ increase in NO₂ exposure. Column headers: CHR: chromosome; Position: Chromosomal position based on NCBI human reference genome assembly Build 37. Mapped Gene: UCSC annotated gene; Gene Group: UCSC gene region feature category; regression coefficient; SE: standard error for regression coefficient; *cg24172570 is located 3.8 kbp upstream of *HIBADH*.

Table S4. Look-up in older children for top 25 findings for prenatal NO₂ exposure in relation to methylation in meta-analysis of newborns.

	Meta-analysis of cord blood results (MeDALL + Generation R + CHS + MoBa)								Replication in 4-year-old children of the MeDALL cohorts			the	Replication in 8-year-old children of the MeDALL + EpiGene cohorts (meta-analysis)							
		NO	₂ pregnancy → N	Methylation at birt	th (n=1,	508)			NO ₂ at pregnancy→ Methylation at 4 yrs (n=733)		NO ₂ at 4 yrs → Methylation at 4 yrs (n=689)			NO ₂ at pregnancy → Methylation at 8 yrs (n=786)			NO ₂ at 8 yrs → Methylation at 8 yrs (n=829)			
Chr	Position (build 37)	CpG	Mapped gene	Gene group	Coef	SE	P-value	Direction	Coef	SE	P- value	Coef	SE	P- value	Coef	SE	P- value	Coef	SE	P- value
19	5709149	cg12283362	LONP1 FDR	Body	-0.007	1.40E-03	1.78E-07	-??-**	0.002	0.0012	0.21	0.001	0.0012	0.37	0.000	0.0013	0.95	0.002	0.0019	0.24
7	27561178	cg24172570	HIBADH* ^{FDR}		-0.004	8.00E-04	3.01E-07	?-**	0.001	0.0009	0.16	0.002	0.0012	0.19	0.001	0.0015	0.69	0.003	0.0020	0.18
10	101380289	cg08973675	SLC25A28 FDR	TSS200	0.005	1.10E-03	2.20E-06	++++	0.002	0.0011	0.03	0.001	0.0012	0.47	0.003	0.0012	0.04	0.003	0.0019	0.11
22	40355732	cg17988310	GRAP2	Body	0.004	9.00E-04	5.25E-06	++++	0.001	0.0014	0.43	0.002	0.0018	0.24	-0.001	0.0017	0.69	0.003	0.0026	0.22
20	61427684	cg14582546	C20orf20	TSS200	0.005	1.10E-03	5.50E-06	++++	-0.001	0.0013	0.30	-0.001	0.0011	0.30	0.001	0.0010	0.44	0.001	0.0016	0.39
22	39323510	cg12276768	APOBEC3A*		0.003	6.00E-04	5.60E-06	++++	0.000	0.0009	0.77	0.001	0.0009	0.22	0.002	0.0010	0.04	0.002	0.0014	0.16
6	30688588	cg21660604	TUBB	Body	0.002	3.00E-04	8.36E-06	++++	0.000	0.0004	0.45	0.000	0.0004	0.86	0.000	0.0004	0.84	0.001	0.0007	0.06
5	77284206	cg26815688	AP3B1*		-0.002	5.00E-04	9.03E-06		0.000	0.0006	0.41	0.001	0.0006	0.16	0.001	0.0006	0.27	-0.001	0.0009	0.40
6	30524763	cg03860665	PRR3;GNL1	5'UTR;1stExon	0.002	5.00E-04	9.17E-06	++++	0.001	0.0007	0.29	0.000	0.0007	0.68	0.000	0.0009	0.63	0.001	0.0011	0.57
7	117824040	cg08301459	NAA38	TSS200	0.002	3.00E-04	9.58E-06	++?+	0.001	0.0005	0.12	0.001	0.0006	0.07	0.000	0.0005	0.75	0.000	0.0007	0.62
6	33359817	cg04757012	KIFC1	Body	0.001	3.00E-04	1.01E-05	++++	0.000	0.0004	0.76	0.000	0.0005	0.82	0.000	0.0004	0.56	0.000	0.0006	0.86
11	74871202	cg12537437	SLCO2B1	Body;5'UTR	-0.004	8.00E-04	1.02E-05	+	-0.001	0.0007	0.34	0.000	0.0007	0.86	0.001	0.0009	0.40	0.001	0.0013	0.47
1	35226135	cg01828548	GJB4	5'UTR	-0.005	1.10E-03	1.08E-05	+	0.001	0.0012	0.22	0.001	0.0015	0.40	-0.004	0.0015	0.01	-0.006	0.0023	0.01
21	46032086	cg26386968	C21orf29; KRTAP10-8	Body;1stExon	-0.007	1.50E-03	1.15E-05	?-	-0.003	0.0011	0.02	-0.002	0.0013	0.11	-0.002	0.0014	0.11	-0.002	0.0022	0.27
9	139607421	cg12657416	FAM69B	Body	0.103	2.36E-02	1.22E-05	?+?+	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
11	34460856	cg03728580	CAT	Body	0.003	7.00E-04	1.43E-05	++++	0.000	0.0010	1.00	0.000	0.0012	0.85	0.000	0.0010	0.97	0.001	0.0016	0.68
2	231809697	cg21022949	GPR55*		0.001	2.00E-04	1.51E-05	++++	0.000	0.0003	0.77	0.000	0.0004	0.68	0.001	0.0004	0.001	0.000	0.0006	0.89
2	98409069	cg06840305	TMEM131	Body	-0.002	4.00E-04	1.51E-05		0.000	0.0006	0.70	0.001	0.0006	0.10	0.000	0.0008	0.84	0.002	0.0011	0.04
12	120967065	cg11075121	COQ5	TSS200	0.002	4.00E-04	1.66E-05	++++	0.000	0.0006	0.48	0.001	0.0007	0.40	0.001	0.0006	0.06	0.002	0.0009	0.03
8	48099615	cg03271173	IGLV8OR8-1*		-0.003	6.00E-04	1.70E-05	+-	-0.001	0.0005	0.33	0.000	0.0007	0.89	0.000	0.0007	0.58	0.000	0.0010	0.75
8	110346503	cg25407888	ENY2;NUDCD1	TSS200	0.003	6.00E-04	1.98E-05	++++	0.000	0.0009	0.70	0.000	0.0009	0.58	0.000	0.0009	0.74	0.000	0.0011	0.73
21	45753677	cg24316255	C21orf2	Body	-0.003	6.00E-04	2.00E-05	+	-0.001	0.0007	0.14	0.000	0.0009	1.00	-0.001	0.0009	0.12	-0.001	0.0012	0.57
17	78851213	cg08314949	RPTOR	Body;Body	0.013	3.10E-03		+-?+	0.015	0.0039	0.0001	0.010	0.0047	0.04	-0.006	0.0053	0.23	0.003	0.0072	0.66
15	59063272	cg01889112	FAM63B	TSS200;TSS200	0.002	4.00E-04		++++	0.000	0.0006	0.89	0.001	0.0007	0.40	0.000	0.0006	0.75	0.001	0.0009	0.45
6	31382102	cg26504614	MICA	Body	-0.005	1.10E-03	2.59E-05	-?	0.000	0.0010	0.65	0.000	0.0010	0.97	-0.003	0.0010	0.001	-0.001	0.0015	0.61

Shown are top 25 CpGs from the discovery meta-analysis ordered by p-value; Estimates are presented per $10 \mu g/m^3$ increase in NO_2 exposure. Chr: chromosome; Direction: Direction of effect across cohorts included in the meta-analysis (MeDALL, Generation R, CHS and MoBa): NO_2 exposure during pregnancy associated with increased (+) or decreased (-) methylation, or missing (?) result; NA=not available.

* cg24172570 is located 3.8 kbp upstream of *HIBADH*, *cg12276768* – 25.2 kbp upstream of *APOBEC3A*, cg26815688 - 21.1 kbp upstream of *AP3B1*, cg21022949 – 19.7 kbp downstream of *GPR55*, cg03271173 - 14.5 kbp upstream of *IGLV80R8-1*.

^{**} Data on methylation of cg12283362 was available in 473 individuals, cg24172570 - in 1282 individuals.

Table S5. Look-up in 4-year-old children with paired cord blood and 4-year-old samples for top 25 findings for prenatal NO₂ exposure in relation to methylation in meta-analysis of newborns.

		Meta-analysis			ion in 4-year- he MeDALL (old children of cohorts					
			NO ₂ pregnancy → Metl	ıylation at birth (ı	n=1,508)					egnancy→ Mo INMA+EDE	ethylation at 4 N (n=277)
Chr	Position (build 37)	СрG	Mapped gene	Gene group	Coef	SE	P-value	Direction	Coef	SE	P-value
19	5709149	cg12283362	LONP1 FDR	Body	-0.007	1.40E-03	1.78E-07	-??-**	0.001	1.58E-03	0.42
7	27561178	cg24172570	HIBADH* FDR		-0.004	8.00E-04	3.01E-07	?-**	0.001	1.28E-03	0.54
10	101380289	cg08973675	SLC25A28 FDR	TSS200	0.005	1.10E-03	2.20E-06	++++	0.004	1.46E-03	0.005
22	40355732	cg17988310	GRAP2	Body	0.004	9.00E-04	5.25E-06	++++	0.005	2.03E-03	0.02
20	61427684	cg14582546	C20orf20	TSS200	0.005	1.10E-03	5.50E-06	++++	0.001	2.02E-03	0.61
22	39323510	cg12276768	++++	0.001	9.76E-04	0.45					
6	30688588	cg21660604	++++	0.000	5.56E-04	0.55					
5	77284206	cg26815688	<i>AP3B1*</i>		-0.002	5.00E-04	9.03E-06		0.000	8.77E-04	0.66
6	30524763	cg03860665	PRR3;GNL1	5'UTR;1stExon	0.002	5.00E-04	9.17E-06	++++	0.001	8.84E-04	0.37
7	117824040	cg08301459	NAA38	TSS200	0.002	3.00E-04	9.58E-06	++?+	0.002	6.55E-04	0.001
6	33359817	cg04757012	KIFC1	Body	0.001	3.00E-04	1.01E-05	++++	0.001	5.24E-04	0.05
11	74871202	cg12537437	SLCO2B1	Body;5'UTR	-0.004	8.00E-04	1.02E-05	+	-0.001	9.14E-04	0.11
1	35226135	cg01828548	GJB4	5'UTR	-0.005	1.10E-03	1.08E-05	+	-0.002	1.57E-03	0.23
21	46032086	cg26386968	C21orf29;KRTAP10-8	Body;1stExon	-0.007	1.50E-03	1.15E-05	?-	-0.006	1.53E-03	0.00005
9	139607421	cg12657416	FAM69B	Body	0.103	2.36E-02	1.22E-05	?+?+	NA	NA	NA
11	34460856	cg03728580	CAT	Body	0.003	7.00E-04	1.43E-05	++++	0.000	1.41E-03	0.77
2	231809697	cg21022949	GPR55*		0.001	2.00E-04	1.51E-05	++++	0.000	4.22E-04	0.95
2	98409069	cg06840305	TMEM131	Body	-0.002	4.00E-04	1.51E-05		-0.001	9.01E-04	0.33
12	120967065	cg11075121	COQ5	TSS200	0.002	4.00E-04	1.66E-05	++++	0.001	8.22E-04	0.14
8	48099615	cg03271173	IGLV8OR8-1*		-0.003	6.00E-04	1.70E-05	+-	-0.001	8.26E-04	0.10
8	110346503 cg25407888 <i>ENY2;NUDCD1</i> TSS200 0.003 6.00E-04 1.98E-05 ++									1.40E-03	0.81
21	45753677	cg24316255	C21orf2	Body	-0.003	6.00E-04	2.00E-05	+	-0.002	1.07E-03	0.15
17	78851213	cg08314949	RPTOR	Body;Body	0.013	3.10E-03	2.06E-05	+-?+	0.017	4.41E-03	0.0002
15	59063272	cg01889112	FAM63B	TSS200;TSS200	0.002	4.00E-04	2.29E-05	++++	0.001	8.21E-04	0.19
6	31382102	cg26504614	MICA	Body	-0.005	1.10E-03	2.59E-05	-?	0.000	1.23E-03	0.81

Shown are top 25 CpGs from the discovery meta-analysis ordered by p-value; Estimates are presented per 10 µg/m³ increase in NO₂ exposure. Column headers: CHR: chromosome; Position: Chromosomal position based on NCBI human reference genome assembly Build 37. Mapped Gene: UCSC annotated gene; Gene Group: UCSC gene region feature category; regression coefficient; SE: standard error for regression coefficient; Direction: Direction of effect across cohorts included in the statistical model (MeDALL, Generation R, CHS and MoBa): NO₂ exposure during pregnancy associated with increased (+) or decreased (-) methylation, or missing (?) result. NA=not available

* cg24172570 is located 3.8 kbp upstream of *HIBADH*, *cg12276768* – 25.2 kbp upstream of *APOBEC3A*, cg26815688 - 21.1 kbp upstream of *AP3B1*, cg21022949 – 19.7 kbp downstream of *GPR55*, cg03271173 - *14.5* kbp upstream of *IGLV80R8-1*.

^{**} Data on methylation of cg12283362 was available in 473 individuals, cg24172570 - in 1282 individuals.

Table S6. Nominally significant CpGs within oxidative stress genes extracted from the epigenome-wide meta-analysis of the association between prenatal NO₂ exposure and newborn cord blood DNA methylation (n=1,508 newborns from MeDALL, Generation R, CHS and MoBa cohorts).

Chr	Position (build 37)	CpG	Mapped gene	Gene group	Coef	SE	P-value	Direction
11	34460856	cg03728580	CAT FDR	Body	0.003	0.001	0.00001	++++
11	34461028	cg17034036	CAT CAT FDR	Body	0.003	0.001	0.0001	++++
2	1482597	cg01385533	TPO FDR	Body	-0.003	0.001	0.0004	-?
1	226023590	cg05935800	EPHX1	Body	-0.002	0.001	0.002	
20	33539306	cg13607138	GSS	Body	-0.003	0.001	0.003	?-
8	107642385	cg17526936	OXR1	Body	-0.002	0.001	0.004	?-
2	1544120	cg19407717	TPO	Body	-0.002	0.001	0.004	
2	1479523	cg13703866	TPO	Body	-0.001	0.000	0.005	
11	34460336	cg07768201	CAT	TSS200	0.003	0.001	0.006	++++
1	226012507	cg03337430	EPHX1	TSS1500;5'UTR	0.001	0.000	0.006	+-++
2	1416855	cg08946720	TPO	TSS1500	-0.002	0.001	0.006	?+
2	1743009	cg01821226	PXDN	Body	-0.001	0.001	0.007	
2	1654326	cg08380973	PXDN	Body	-0.003	0.001	0.009	
2	1518383	cg24215279	TPO	Body	0.001	0.000	0.011	+-+-
6	160113813	cg14515483	SOD2	Body	0.001	0.000	0.012	++++
2	1652518	cg26063629	PXDN	Body	-0.002	0.001	0.012	+
11	34460351	cg03720043	CAT	TSS200	0.003	0.001	0.012	++++
11	67351273	cg26250609	GSTP1	1stExon;5'UTR	0.002	0.001	0.013	++++
17	26120702	cg01396112	NOS2	Body	0.000	0.000	0.016	+
2	1488516	cg02892893	TPO	Body	-0.001	0.001	0.018	?-
15	45406362	cg10778736	DUOX2	TSS200	0.002	0.001	0.018	++++
15	45405370	cg07821960	DUOX2	Body;TSS1500	0.001	0.000	0.018	++++
17	26084080	cg15088880	NOS2	3'UTR	0.005	0.002	0.019	??+-
8	52618305	ch.8.1157478F	PXDNL	Body	0.004	0.002	0.020	++?+
2	1425205	cg06972972	TPO	Body	-0.001	0.000	0.021	
2	201450731	cg12627583	AOX1	1stExon;5'UTR	0.001	0.001	0.021	+++-
7	2289888	cg13748354	NUDT1	Body	0.038	0.016	0.022	?+?+
2	1478219	cg25013910	TPO	Body	-0.004	0.002	0.022	?-
11	67350499	cg05244766	GSTP1	TSS1500	-0.002	0.001	0.024	-?-+
3	38207809	cg15789250	OXSR1	Body	0.005	0.002	0.025	++++
2	201450690	cg02144933	AOX1	TSS200	0.001	0.000	0.029	++++
16	69760301	cg06790860	NQO1	Body	0.001	0.000	0.032	+-++
2	1668758	cg12624031	PXDN	Body	-0.001	0.001	0.034	
17	56274480	cg08105265	EPX	Body	0.002	0.001	0.034	+?++
8	52721657	cg02090805	PXDNL	Body	0.001	0.001	0.035	+++-
8	107738435	cg12114888	OXR1	Body	0.005	0.002	0.037	-+++
2	1682068	cg09996777	PXDN	Body	-0.003	0.001	0.040	?-
2	1493713	cg06173919	TPO	Body	-0.002	0.001	0.041	?+

1	225996810	cg21826272	EPHX1	TSS1500	-0.002	0.001	0.043	?-
7	2281541	cg04305677	NUDT1	TSS1500	0.000	0.000	0.044	++
11	67350491	cg08925882	GSTP1	TSS1500	-0.004	0.002	0.045	??
2	178128234	cg16842060	NFE2L2	Body;1stExon;5'UTR	0.000	0.000	0.047	+-++
7	2284600	cg03663120	NUDT1	Body	0.007	0.004	0.048	++?+
1	53068579	cg23272399	GPX7	Body	-0.001	0.001	0.049	
6	160114681	cg04311230	SOD2	TSS1500	0.000	0.000	0.049	-+++

Shown here are the 45 nominally significant (p<0.05) CpGs ordered by p-value. Three CpGs were statistically significant using genome-wide significance threshold (FDR p<0.05). Results presented per 10 μ g/m³ increase in prenatal NO₂ exposure.

Column headers: CHR=chromosome; Position=Chromosomal position based on NCBI human reference genome assembly Build 37. Mapped Gene=UCSC annotated gene; Gene Group=UCSC gene region feature category; regression coefficient; SE=standard error for regression coefficient; Direction=Direction of effect across cohorts included in the statistical model (MeDALL, Generation R, CHS and MoBa): NO₂ exposure during pregnancy associated with increased (+) or decreased (-) methylation, or missing (?) result.

Table S7. All available CpGs mapped to *CAT* gene from the epigenome-wide meta-analysis of the association between NO₂ exposure during pregnancy and newborn cord blood DNA methylation (N=1,508 newborns from MeDALL, Generation R, CHS and MoBa cohorts).

Chr	Position (build 37)	CpG	Mapped gene	Gene group	Coef	SE	P-value	Direction
11	34460856	cg03728580	CAT	Island	0.003	7.00E-04	0.00001	++++
11	34461028	cg17034036	CAT	S_Shore	0.002	6.00E-04	0.0001	++++
11	34460336	cg07768201	CAT	Island	0.003	1.10E-03	0.01	++++
11	34460351	cg03720043	CAT	Island	0.003	1.00E-03	0.01	++++
11	34460516	cg06908474	CAT	Island	0.002	1.00E-03	0.07	++++
11	34460386	cg02109652	CAT	Island	0.002	1.00E-03	0.12	-+++
11	34460107	cg20234170	CAT	N_Shore	0.001	8.00E-04	0.19	++-+
11	34460557	cg01847719	CAT	Island	0.002	1.50E-03	0.22	++++
11	34460182	cg22159421	CAT	Island	0.001	6.00E-04	0.23	+-++
11	34460318	cg06027906	CAT	Island	0.000	3.00E-04	0.24	++++
11	34460172	cg24099074	CAT	Island	0.000	2.00E-04	0.25	++++
11	34460298	cg20731136	CAT	Island	0.001	4.00E-04	0.26	+-++
11	34464477	cg09106728	CAT	S_Shelf	-0.001	1.60E-03	0.48	?+
11	34492984	cg17098995	CAT	OpenSea	0.000	3.00E-04	0.82	+-
11	34460789	cg14316565	CAT	Island	0.000	6.00E-04	0.96	+-++

Shown are all CpGs within *CAT* gene ordered by p-value; All results presented per 10 μg/m³ increase in prenatal NO₂ exposure. Column headers: CHR: chromosome; Mapped Gene: UCSC annotated gene; Gene Group: UCSC gene region feature category; Coef: regression coefficient; Direction: Direction of effect across cohorts included in the statistical model (MeDALL, Generation R, CHS and MoBa): NO₂ exposure during pregnancy associated with increased (+) or decreased (-) methylation, or missing (?) result.

Table S8. All available CpGs mapped to *TPO* gene from the epigenome-wide meta-analysis of the association between NO₂ exposure during pregnancy and newborn cord blood DNA methylation (N=1,508 newborns from MeDALL, Generation R, CHS and MoBa cohorts).

Chr	Position (build 37)	CpG	Mapped gene	Gene group	Coef	SE	P-value	Direction
2	1482597	cg01385533	TPO	S_Shore	-0.003	8.00E-04	0.0004	-?
2	1544120	cg19407717	TPO	N_Shore	-0.002	8.00E-04	0.004	
2	1479523	cg13703866	TPO	N_Shore	-0.001	3.00E-04	0.005	
2	1416855	cg08946720	TPO	OpenSea	-0.002	8.00E-04	0.01	?+
2	1518383	cg24215279	TPO	S_Shelf	0.001	2.00E-04	0.01	+-+-
2	1488516	cg02892893	TPO	OpenSea	-0.001	5.00E-04	0.02	?-
2	1425205	cg06972972	TPO	OpenSea	-0.001	4.00E-04	0.02	
2	1478219	cg25013910	TPO	N_Shelf	-0.004	1.90E-03	0.02	?-
2	1493713	cg06173919	TPO	Island	-0.002	9.00E-04	0.04	?+
2	1482385	cg26140366	TPO	S_Shore	-0.002	9.00E-04	0.05	+
2	1417153	cg12680131	TPO	OpenSea	-0.002	1.40E-03	0.09	
2	1544321	cg04770020	TPO	Island	0.000	2.00E-04	0.11	+
2	1494063	cg01681351	TPO	Island	-0.003	1.70E-03	0.11	??
2	1500117	cg05678658	TPO	OpenSea	-0.002	1.20E-03	0.11	+
2	1426181	cg03347837	TPO	OpenSea	-0.001	7.00E-04	0.12	-+-+
2	1493387	cg00626390	TPO	N_Shore	-0.001	6.00E-04	0.13	?+
2	1482838	cg12666976	TPO	S_Shore	-0.002	1.50E-03	0.13	-++-
2	1497815	cg13414059	TPO	S_Shelf	-0.002	1.00E-03	0.16	-+
2	1417109	cg16016036	TPO	OpenSea	-0.001	7.00E-04	0.16	-+
2	1516247	cg07713008	TPO	Island	-0.001	1.00E-03	0.16	
2	1494263	cg23596425	TPO	S_Shore	-0.003	1.90E-03	0.17	++
2	1416889	cg00040862	TPO	OpenSea	0.001	8.00E-04	0.20	-++-
2	1452665	cg19368625	TPO	OpenSea	-0.001	1.10E-03	0.21	++
2	1426094	cg14108581	TPO	OpenSea	-0.001	1.00E-03	0.21	?-
2	1417431	cg10370591	TPO	OpenSea	0.001	6.00E-04	0.21	++++
2	1482915	cg23985797	TPO	S_Shore	0.001	9.00E-04	0.21	+-++
2	1507739	cg10672136	TPO	OpenSea	-0.001	1.20E-03	0.24	++
2	1488252	cg09337427	TPO	OpenSea	0.001	7.00E-04	0.24	+-+-
2	1544370	cg18995558	TPO	Island	-0.001	6.00E-04	0.25	
2	1426068	cg10531568	TPO	OpenSea	-0.001	7.00E-04	0.26	-++-
2	1507694	cg15375772	TPO	OpenSea	-0.002	1.70E-03	0.27	++
2	1498138	cg03545077	TPO	S_Shelf	0.001	1.30E-03	0.27	++?+
2	1498065	cg18019372	TPO	S_Shelf	-0.002	1.60E-03	0.27	+
2	1418073	cg23136645	TPO	OpenSea	0.000	4.00E-04	0.29	-++-
2	1418028	cg09757588	TPO	OpenSea	0.000	4.00E-04	0.31	+
2	1425319	cg02963613	TPO	OpenSea	-0.001	7.00E-04	0.33	+-
2	1487936	cg19905414	TPO	OpenSea	-0.001	8.00E-04	0.34	?-
2	1484593	cg08025464	TPO	S_Shelf	-0.003	2.90E-03	0.35	-+
2	1544474	cg06697522	TPO	Island	-0.001	7.00E-04	0.39	+
2	1544076	cg17518079	TPO	N_Shore	0.001	7.00E-04	0.39	++-+

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2	1426328	cg14691886	TPO	OpenSea	-0.001	6.00E-04	0.40	-+?-
2	1482963	cg20957776	TPO	S_Shore	-0.001	1.30E-03	0.40	++
2	1488175	cg17299636	TPO	OpenSea	-0.001	1.00E-03	0.43	?-
2	1479763	cg02332115	TPO	N_Shore	-0.001	1.10E-03	0.44	
2	1500255	cg21759048	TPO	OpenSea	-0.001	8.00E-04	0.44	+-
2	1461560	cg15301057	TPO	OpenSea	-0.001	8.00E-04	0.44	-+?-
2	1543221	cg05090359	TPO	N_Shore	0.001	6.00E-04	0.45	+++-
2	1484125	cg21856680	TPO	S_Shelf	0.001	6.00E-04	0.46	+-+-
2	1513998	cg03133821	TPO	N_Shelf	0.000	6.00E-04	0.46	-+
2	1543327	cg04319651	TPO	N_Shore	0.000	6.00E-04	0.47	+
2	1452260	cg16407924	TPO	OpenSea	-0.001	1.90E-03	0.48	+-?-
2	1479810	cg01112527	TPO	N_Shore	-0.001	7.00E-04	0.48	
2	1517047	cg14926103	TPO	S_Shore	0.001	7.00E-04	0.50	++
2	1494681	cg14011419	TPO	S_Shore	0.001	1.10E-03	0.52	++?-
2	1500368	cg17494408	TPO	OpenSea	0.000	7.00E-04	0.54	-++-
2	1489909	cg25250661	TPO	N_Shelf	-0.001	1.30E-03	0.56	+
2	1417164	cg06500727	TPO	OpenSea	-0.001	9.00E-04	0.57	++
2	1516240	cg24798914	TPO	Island	0.000	4.00E-04	0.58	+
2	1497906	cg02924495	TPO	S_Shelf	0.000	7.00E-04	0.59	++?+
2	1544352	cg06574769	TPO	Island	0.000	7.00E-04	0.60	++
2	1497868	cg26541429	TPO	S_Shelf	0.000	7.00E-04	0.61	+-
2	1488565	cg04658693	TPO	OpenSea	0.000	7.00E-04	0.61	-++-
2	1418294	cg15977002	TPO	OpenSea	0.000	7.00E-04	0.68	+
2	1426845	cg07088935	TPO	OpenSea	0.000	1.00E-03	0.68	++
2	1542097	cg01028140	TPO	N_Shelf	0.000	9.00E-04	0.69	++
2	1507832	cg23239637	TPO	OpenSea	0.000	2.00E-04	0.71	+-
2	1543996	cg11275685	TPO	N_Shore	0.000	6.00E-04	0.72	++
2	1481097	cg14601038	TPO	Island	-0.001	2.10E-03	0.72	++
2	1480789	cg07932899	TPO	Island	-0.001	1.80E-03	0.73	-+++
2	1487895	cg19585646	TPO	OpenSea	0.000	1.00E-03	0.73	++?-
2	1488638	cg14935163	TPO	OpenSea	0.000	3.00E-04	0.76	++
2	1425035	cg14478255	TPO	OpenSea	0.000	7.00E-04	0.77	-+-+
2	1425560	cg26277787	TPO	OpenSea	0.000	7.00E-04	0.80	++
2	1481492	cg08537127	TPO	Island	0.001	5.50E-03	0.81	?-?+
2	1417248	cg07083862	TPO	OpenSea	0.000	8.00E-04	0.86	+-+-
2	1543546	cg21913853	TPO	N_Shore	0.000	5.00E-04	0.87	++
2	1452367	cg26605809	TPO	OpenSea	0.000	1.30E-03	0.88	-+++
2	1482185	cg21997141	TPO	S_Shore	0.000	8.00E-04	0.89	+-
2	1426301	cg05544715	TPO	OpenSea	0.000	3.00E-04	0.92	-+
2	1426674	cg24216893	TPO	OpenSea	0.000	8.00E-04	0.92	+
2	1516061	cg01312658	TPO	N_Shore	0.000	5.00E-04	0.96	+-
2	1480944	cg14793137	TPO	Island	0.000	2.80E-03	0.97	-+++
2	1415911	cg25349955	TPO	OpenSea	0.000	6.00E-04	0.97	++
2	1425620	cg03795587	TPO	OpenSea	0.000	6.00E-04	0.98	++
2	1516352	cg13661012	TPO	Island	0.000	4.00E-04	0.99	+-+-
2	1488314	cg04152326	TPO	OpenSea	0.000	5.00E-04	1.00	++

Shown are all CpGs within TPO gene ordered by p-value, with exception of cg01957222 only available in MoBa cohort. All results presented per 10 μ g/m³ increase in prenatal NO₂ exposure.

Column headers: CHR: chromosome; Mapped Gene: UCSC annotated gene; Gene Group: UCSC gene region feature category; Coef: regression coefficient; Direction: Direction of effect across cohorts included in the statistical model (MeDALL, Generation R, CHS and MoBa): NO₂ exposure during pregnancy associated with increased (+) or decreased (-) methylation, or missing (?) result.

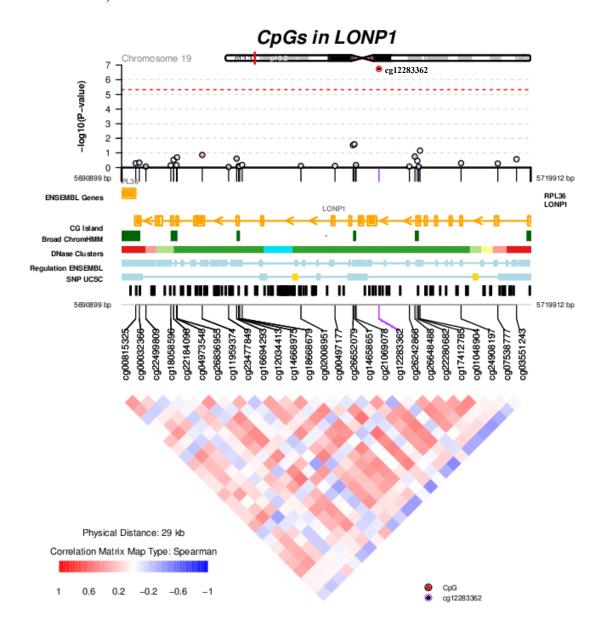
Table S9. Look-up in older children for top CpGs in the CAT and TPO genes for prenatal NO₂ exposure in relation to methylation in meta-analysis of newborns (n=1,508).

N	leta-analysi	s of cord bloo	d results (MeDAL	L+ Ge	neration R	+ CHS +	MoBa)	Replication in 4-year-old children of the MeDALL cohorts						Replication in 8-year-old children of the MeDALL + EpiGene cohorts (meta-analysis)						
NO ₂ pregnancy → Methylation at birth (n=1,508)										NO ₂ at pregnancy→ Methylation at 4 yrs (n=733)			NO ₂ at 4 yrs → Methylation at 4 yrs (n=689)			NO ₂ at pregnancy → Methylation at 8 yrs (n=786)			NO ₂ at 8 yrs → Methylation at 8 yrs (n=829)		
	Position (build		Mapped	Gene							P-			P-			P-		`	P-	
Chr	37)	CpG	gene	group	Coef	SE	P-value	Direction	Coef	SE	value	Coef	SE	value	Coef	SE	value	Coef	SE	value	
11	34460856	cg03728580	CAT FDR	Body	0.003	7.00E-04	0.00001	++++	0.0000	9.62E-04	1.00	0.000	1.23E-03	0.85	0.0000	1.00E-03	0.97	0.0006	1.60E-03	0.68	
11	34461028	cg17034036	CAT^{FDR}	Body	0.002	6.00E-04	0.0001	++++	0.0003	7.78E-04	0.72	-0.0001	8.47E-04	0.93	0.0002	1.00E-03	0.82	-0.0003	1.20E-03	0.80	
2	1482597	cg01385533	TPO FDR	Body	-0.003	8.00E-04	0.0004	-?	-0.0006	7.31E-04	0.40	-0.002	7.87E-04	0.04	-0.0018	8.00E-04	0.04	-0.0016	1.30E-03	0.22	

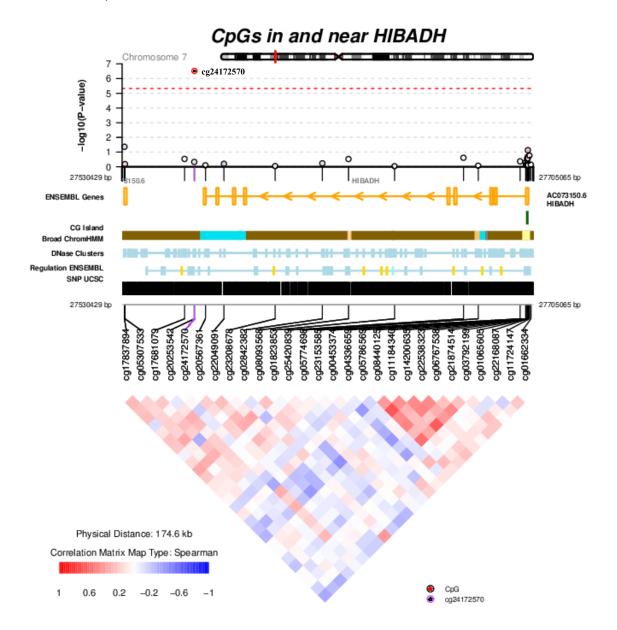
Shown are top CpGs within the *CAT* and *TPO* genes from the discovery meta-analysis ordered by p-value; Estimates are presented per 10 µg/m³ increase in NO₂ exposure. Column headers: CHR: chromosome; Position: Chromosomal position based on NCBI human reference genome assembly Build 37. Mapped Gene: UCSC annotated gene; Gene Group: UCSC gene region feature category; regression coefficient; SE: standard error for regression coefficient; Direction: Direction of effect across cohorts included in the statistical model (MeDALL, Generation R, CHS and MoBa): NO₂ exposure associated with increased (+) or decreased (-) methylation, or missing (?) result.

Figure S3

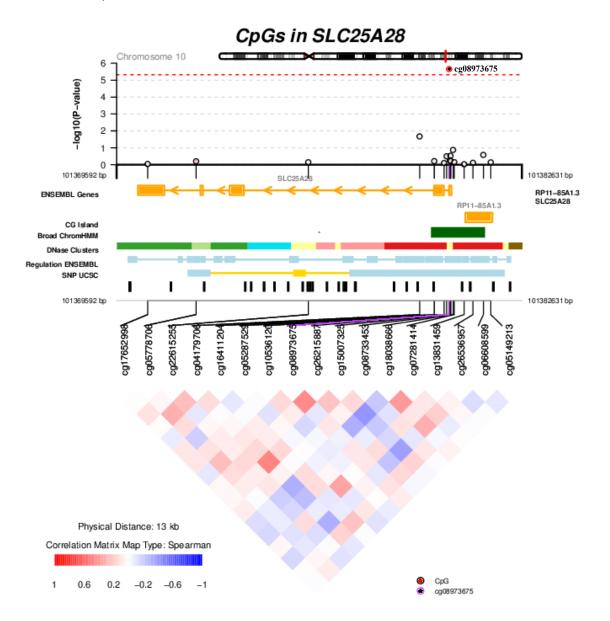
A)



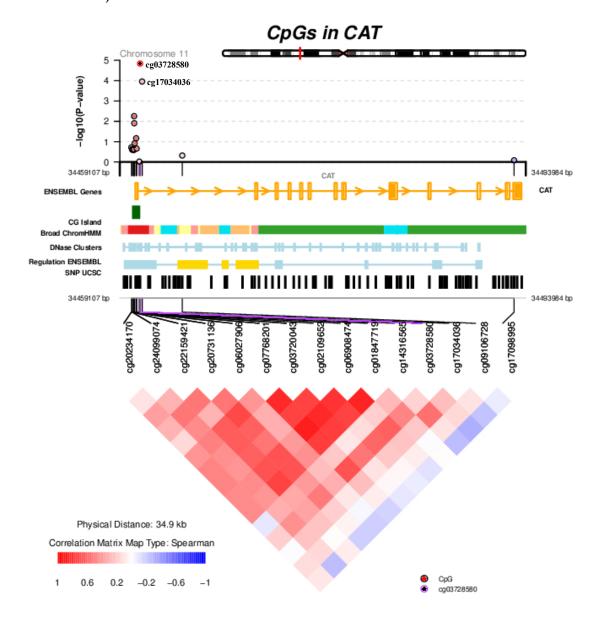
B)



C)



D)



E)

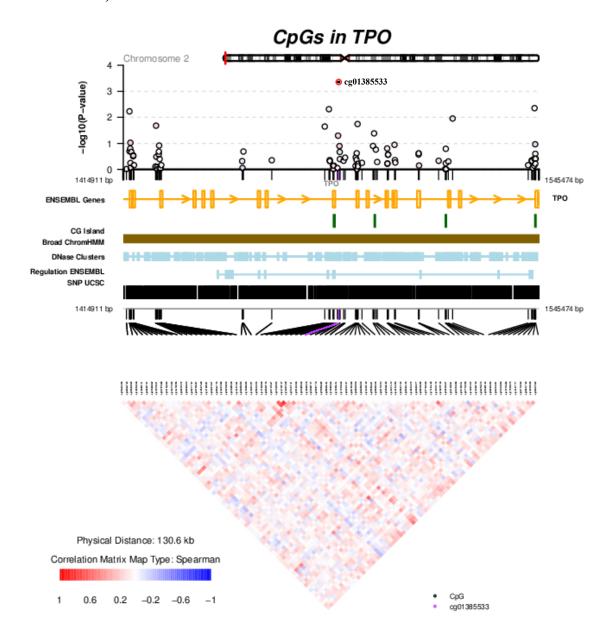


Figure S3. Regional plots and co-methylation patterns for the genes containing FDR-significant EWAS CpG sites (circles): A) *LONP1* (cg12283362), B) *HIBADH* (cg24172570), C) *SLC25A28* (cg08973675), as well as D) *CAT* (cg03728580 and cg17034036) and E) *TPO* (cg01385533). -log10(p values) from the meta-analysis, CpGs indicated by dots, color coded based on pairwise correlation with neighboring CpGs. The lower panel demonstrate pairwise correlation matrix across the displayed CpGs.

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References

Aryee MJ, Jaffe AE, Corrada-Bravo H, et al. 2014. Minfi: A flexible and comprehensive bioconductor package for the analysis of infinium DNA methylation microarrays. Bioinformatics 30:1363-1369.

Beelen R, Hoek G, Vienneau D, et al. 2013. Development of no2 and nox land use regression models for estimating air pollution exposure in 36 study areas in europe - the escape project. Atmos Environ 72:10-23.

Bolstad BM, Irizarry RA, Astrand M, et al. 2003. A comparison of normalization methods for high density oligonucleotide array data based on variance and bias. Bioinformatics 19:185-193.

Bousquet J, Anto J, Auffray C, et al. 2011. Medall (mechanisms of the development of allergy): An integrated approach from phenotypes to systems medicine. Allergy 66:596-604.

Chen YA, Lemire M, Choufani S, et al. 2013. Discovery of cross-reactive probes and polymorphic cpgs in the illumina infinium humanmethylation 450 microarray. Epigenetics 8:203-209.

Guxens M, Ballester F, Espada M, et al. 2012. Cohort profile: The inma--infancia y medio ambiente--(environment and childhood) project. Int J Epidemiol 41:930-940.

Hannam K, McNamee R, De Vocht F, et al. 2013. A comparison of population air pollution exposure estimation techniques with personal exposure estimates in a pregnant cohort. Environ Sci Process Impacts 15:1562-1572.

Heude B, Forhan A, Slama R, et al. 2015. Cohort profile: The eden mother-child cohort on the prenatal and early postnatal determinants of child health and development. Int J Epidemiol.

Jaddoe VW, van Duijn CM, Franco OH, et al. 2012. The generation r study: Design and cohort update 2012. Eur J Epidemiol 27:739-756.

Joubert B, Felix J, Yousefi P, et al. 2016. DNA methylation in newborns and maternal smoking in pregnancy: Genome-wide consortium meta-analysis. Am J Hum Genet 98(4):680-696.

Kruithof CJ, Kooijman MN, van Duijn CM, et al. 2014. The generation r study: Biobank update 2015. Eur J Epidemiol 29:911-927.

Magnus P, Irgens LM, Haug K, et al. 2006. Cohort profile: The norwegian mother and child cohort study (moba). Int J Epidemiol 35:1146-1150.

McConnell R, Berhane K, Yao L, et al. 2006. Traffic, susceptibility, and childhood asthma. Environ Health Perspect 114:766-772.

Noushmehr H, Weisenberger DJ, Diefes K, et al. 2010. Identification of a cpg island methylator phenotype that defines a distinct subgroup of glioma. Cancer Cell 17:510-522.

Pedersen M, Giorgis-Allemand L, Bernard C, et al. 2013. Ambient air pollution and low birthweight: A european cohort study (escape). The lancet Respiratory medicine 1:695-704.

Peters JM, Avol E, Gauderman WJ, et al. 1999a. A study of twelve southern california communities with differing levels and types of air pollution. Ii. Effects on pulmonary function. Am J Respir Crit Care Med 159:768-775.

Peters JM, Avol E, Navidi W, et al. 1999b. A study of twelve southern california communities with differing levels and types of air pollution. I. Prevalence of respiratory morbidity. Am J Respir Crit Care Med 159:760-767.

Pidsley R, CC YW, Volta M, et al. 2013. A data-driven approach to preprocessing illumina 450k methylation array data. BMC Genomics 14:293.

Rivera-Gonzalez LO, Zhang Z, Sanchez BN, et al. 2015. An assessment of air pollutant exposure methods in mexico city, mexico. J Air Waste Manag Assoc 65:581-591.

Ronningen KS, Paltiel L, Meltzer HM, et al. 2006. The biobank of the norwegian mother and child cohort study: A resource for the next 100 years. Eur J Epidemiol 21:619-625.

Thacher JD, Gruzieva O, Pershagen G, et al. 2016. Parental smoking and development of allergic sensitization from birth to adolescence. Allergy 71:239-248.

Touleimat N, Tost J. 2012. Complete pipeline for infinium((r)) human methylation 450k beadchip data processing using subset quantile normalization for accurate DNA methylation estimation. Epigenomics 4:325-341.

Triche TJ, Jr., Weisenberger DJ, Van Den Berg D, et al. 2013. Low-level processing of illumina infinium DNA methylation beadarrays. Nucleic Acids Res 41:e90.

Van den Hooven EH, Pierik FH, Van Ratingen SW, et al. 2012. Air pollution exposure estimation using dispersion modelling and continuous monitoring data in a prospective birth cohort study in the netherlands. Environ Health-Glob 11:9.

Wickman M, Kull I, Pershagen G, et al. 2002. The bamse project: Presentation of a prospective longitudinal birth cohort study. Pediatr Allergy Immunol 13 Suppl 15:11-13.